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ANIMAL TRANSACTION POLICY

Updated 2009

The British and Irish Association of Zoos and Aquariums

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Document Status

The BIAZA Animal Transaction Policy was first ratified by the Council on 10th November 1999 and was effective from that date superseding all other existing documents.

*Sections of this policy are mandatory and **MUST** be complied with in full in accordance with Section 5 of The Obligations of Membership and in accordance with the Section 3-(iii) c of The Constitution. The sections of this policy that are mandatory are indicated by the word **MUST**. The remainder of the document should be accepted as Guidelines. Members should comply where possible with these guidelines and in circumstances where this is not possible Members may need to justify their actions.*

The Animal Transaction Policy is under constant review and will be amended, as new and relevant information becomes available. Members have an opportunity to comment on this policy at any time and should lodge such representations or comments with the BIAZA Office.

Furthermore, this document is in the public domain and may be released to person or persons expressing an interest in the Animal Transaction Policy.

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Preamble

BIAZA recognises a strong ethical tradition within the organisation and Member Collections, which forms the basis of an Animal Transaction Policy based on a respect for animals, and a need to maintain viable populations of species. Current standards of animal management, nutrition and veterinary care make responsible zoos producers rather than consumers of wildlife for many species. Therefore, to address this situation, this document lays down a policy for the acquisition and disposition of animals.

All arrangements for transportation of animals ***MUST*** comply with relevant legislation, such as the Welfare of Animals During Transport legislation, the regulations of the International Air Transport Association (IATA), *Live Animals Regulations*, the *WAZA Code of Ethics and Animal Welfare (2003)* (Appendix 6) and the EAZA Conditions of Animal Transfers (Appendix 4).

I. Animal Acquisitions

Animals are acquired by BIAZA collections in three ways:

- (a) by birth;
- (b) by loan, purchase or donation from a third party;
- (c) or from the wild under particular circumstances.

When acquiring animals, BIAZA collections are responsible for ensuring that the source of animals is primarily confined to those born in captivity (but for special cases see Appendix 10) and that this is best achieved by direct zoo to zoo contact. This does not preclude the receipt of animals resulting from confiscation or rescue. It is also recognised that under certain circumstances, there may be a legitimate need for conservation breeding programmes, education programmes and basic biological studies to obtain animals from the wild; though BIAZA collections should obtain animals from the wild only in carefully considered and approved circumstances, normally involving a managed breeding programme. The use of animal dealers should be avoided in most circumstances (see section III).

Strictures applying to animal acquisitions.

1. The receiving collection ***MUST*** be capable of providing the acquired species with appropriate levels of husbandry, taking into account the animal's behavioural and physiological needs.
2. The acquisition ***MUST*** be legal; and ***MUST*** comply with all international, regional and national laws, e.g. CITES Regulations. Members should take care to ensure that animals acquired from animal dealers (and see Section III) have not been laundered through third parties or countries.
3. Acquisition from less than ideal facilities may be considered if the acquisition is of welfare benefit to the animal or if the acquisition contributes to a breeding programme. Collections should ensure, however, that they are not contributing financially or morally to the continuation of such unsatisfactory facilities (*However, this does not preclude the purchase of an animal if there is no other option*).
4. Acquisition from approved certified sustainable sources. For example Marine Aquarium Council (MAC) and some national certified programmes for fish and invertebrates. See Appendix 10

II. Disposal of Surplus Animals

For the purpose of this document, a "surplus" animal is defined as any individual, which a current collection no longer wishes to house, for any reason.

Current standards of animal management, nutrition and veterinary care make responsible zoos producers rather than consumers of wildlife for many species. With lower mortality and longer reproductive lives than those of their wild counterparts, zoo populations can rapidly outgrow available space. Breeding should not continue unchecked unless offspring can be found suitable homes. At the same time, it is recognised that contraception may not always be effective, desirable, practical or commensurate with animal health and welfare. Furthermore reproduction may be considered an integral part of the quality of life and natural behaviour of animals. BIAZA supports those members who recognise the problems caused by irresponsible breeding and disposition of animals (section A.), and adhere to the measures and procedures listed below (sections, C & D).

The disposal of surplus animals **MUST** be legal; it **MUST** comply with all international, regional and national laws, e.g. CITES Regulations and the UK Secretary of State's Standards referring to the disposal of stock, which state: "*Surplus zoo stock only to be passed to persons with the appropriate facilities, resources and expertise conforming with the Five Principles. Precautions should also be taken to ensure that recipients are likely to safeguard the animal's welfare in any subsequent transaction. If animals bred in zoos are sold as pets to the general public, a licence is required from the local authority under the Pet Animals Act 1951*". (N.B. the Pet Animals Act should be eventually incorporated into the Animal Welfare Act 2006).

A. Breeding and disposal of surplus:

BIAZA collections **MUST** breed and dispose of animals in a responsible manner. BIAZA collections **MUST NOT**:

- (a) send animals to zoos or aquariums with inadequate facilities and/or expertise;
- (b) send animals to institutions other than zoos or aquariums which would not normally be considered suitable outlets, which may include circuses, some research institutions, animal dealers, welfare organisations involved in inappropriate reintroductions, etc.,
- (c) Private individuals, unless absolutely certain of their ability to care for the animal???

or disrupt the smooth running of species management initiatives by:

- (a) having animals taking up space needed for higher priority species (or individuals) in the RCP;
- (b) relocating highly inbred/closely related/hybrid specimens to regions not yet equipped to deal with such problems effectively;
- (c) using dealers/brokers whose records are inadequate, which may cause confusion of identities in transit (and see Section III).
- (d) Moving animals in or out of management programmes without the approval of the programme coordinator.

B. Categories of surplus

Surplus individuals may arise through:

- (a) changing status of the taxon in the Regional Collection Plan (RCP);
- (b) exceeding demand due to an inability to predict it;
- (c) exceeding demand due to large litter sizes
- (d) failure of contraceptive measures;
- (e) inappropriate sex ratios;
- (f) late recommendations by species managers;
- (g) breeding against recommendations of species managers;
- (h) inability to prevent breeding;
- (i) desirability to maintain breeding for welfare, training or animal management benefits.

C. Minimising surplus

The following measures may be taken to reduce or prevent the production of surplus:

- (a) adhering to breeding/non-breeding recommendations by species managers;
- (b) planning collections nationally/regionally, thereby increasing the ability to predict demand and to breed accordingly;
- (c) employing appropriate husbandry techniques (Appendix 1);
- (d) employing appropriate veterinary techniques (Appendix 1);
- (e) euthanasia (including culling) (Appendices 1 and 8).

N.B. Efforts should be made to document and publish techniques to limit births for those species in regular surplus, and discussions with those directly involved should be held before the introduction of culling as an institution policy.

D. Disposal of surplus

Release to the wild should not be used a method of disposal of surplus stock. All animal release programmes should comply with IUCN Guidelines for Re-Introduction (1995) and see Hall (2003) for comments on returning large sharks and similar marine animals to the wild.

Members covered by the Zoo Licensing Act (1981) (Amendment) Regulations 2002 **MUST** comply with the Secretary of State's current, relevant Standards referring to the disposal of stock..

Members not covered by the Zoo Licensing Act (1981) (Amendment) Regulations 2002 **MUST** comply with these Standards as a condition of membership, in accordance with Section 5.2 of BIAZA's Constitution.

It is recognised that current "custom and practice" in regard to the disposal of surplus animals may differ between taxa, but in principle the same duty of care exists for all animals.

The dispersing institution should have either inspected the facilities at, or received suitable references concerning, the receiving collection. Such reassurances, regarding facilities and expertise are **MANDATORY** for all species included in Category I of the Hazardous Animals List of the *Secretary of States Standards of Modern Zoo Practice (2000)*, all primates, and any other species that Council may from time to time nominate.

Whilst this high level of scrutiny should be applied for all taxa if possible, it is recognised that for certain taxa a lower level of reassurance may be appropriate. In these cases, members should use the approved BIAZA Proforma (Appendix 2) to establish a minimum level of information prior to implementing the transaction.

For species normally domesticated in the British Isles and part of accepted agricultural transactions, e.g. cattle, the use of the Proforma may be inappropriate and is not mandatory. In these circumstances members should follow the Codes of Recommendations for the Welfare of Livestock, prepared by the Farm Animal Welfare Council (FAWC) and published by the Ministry of Agriculture, Fisheries and Food (MAFF now Defra), (Appendix 3). In the case of other domesticated stock, the Proforma should be used when proposing relocation to another collection, institution or individual.

The following lists the facilities to which animals should be dispersed, and the precautions which should be taken to ensure that the process is completed responsibly:

- (a) species coordinated through a managed breeding programme in Europe, i.e. an EEP or ESBa (Appendix 4), or internationally managed programmes, e.g. International Studbooks, GSMPs;

1. animals bred according to the recommendations of the coordinator should be relocated as

directed.

2. approval should be sought from the coordinator before animals born surplus to the programme are transferred from the collection.

N.B. Responsibility for placing an animal in a suitable facility ultimately rests with the sending institution.

(b) species not coordinated through a managed programme in Europe should first be offered without broker/dealer involvement to;

1. BIAZA Zoos & Aquariums or Accredited Associates (through BIAZA's Available and Wanted List),
2. EAZA Zoos (through the EAZA Available & Wanted List),
3. Coordinators responsible for managing the species outside Europe (if any),
4. EEP/ESB participating but not /EAZA members, through institutional surplus & wanted lists or through EAZA TAGs.
5. Zoos outside Europe associated with a national/regional zoo organisation, and where an animal is placed outside the region it should be according to each region's preferred procedure and where relevant follow the Global Species Management Plan for the taxon.
6. Other appropriate institutions, bearing in mind that it may be preferable to work with a cooperative private facility than with a zoo that declines to work in cooperation with either national or regional zoo associations.

N.B. Where the region is not yet equipped with a regional structure capable of making decisions on the suitability of available stock for that region, the onus is on the sending zoo to behave in a responsible manner, avoiding in particular sending potential breeding pairs of closely related stock or hybrid specimens, without notifying the receiving zoo of the potential problems this may cause.

Euthanasia (including culling)

If, it is deemed necessary to euthanase an animal, the euthanasia technique used ***MUST*** ensure a quick death without suffering. Defra (2003) Circular Guidance (Annex G) provides a useful decision tree to help decide if an animal should be put down, this is reproduced in Appendix 8.

Furthermore, when an animal is euthanased, it is important to provide for the optimal use of the materials available, particularly for studies, which may benefit the species. A post mortem examination should be performed and biological material preserved for research and gene conservation as needed. Tissue collection and post-mortem protocols, and addresses of recommended recipients of material are available from the BIAZA Office or relevant Taxon Working Group and EAZA TAG chairs.

Euthanasia may be practised where the quality of an animal's life is considered to be irreversibly compromised, and may also be considered:

- (a) where the only alternative is permanent transfer to unsuitable accommodation;
- (b) for young animals born despite reproduction-limiting measures or recommendations, that have reached weaning age, or the age at which they would naturally leave the parents or natal group;
- (c) for injured animals, donated or otherwise acquired, that cannot be rehabilitated;
- (d) for hybrids and animals of an unknown or undefined subspecies in cases where this is considered of importance in the context of a managed programme;
- (e) for animals which cannot make a breeding contribution because of old age, genetic over-representation, disease, or the possession of undesirable inheritable or behavioural traits.
- (f) where euthanasia is the only suitable or available measure of population management;
- (g) When, in order to maintain reproductively viable and genetically healthy populations of some

species in captivity, females may need to remain reproductively active to avoid reproductive senescence and or pathology. Furthermore, old or genetically redundant individuals can have a negative effect on the viability of captive populations by taking up valuable and finite space or by compromising effective genetic management. As a result zoos and aquariums may contain individuals which are surplus to the requirements of effective population management. Curtailing reproduction to avoid the generation of such surplus animals may compromise the welfare and reproductive viability of individual females and, ultimately, populations. Humane euthanasia is a legitimate option in these circumstances.

*N.B. In the case of proposed euthanasia, careful consideration and discussion with those directly involved **MUST** be undertaken for each individual animal (and see Appendix 8).*

E. Disposal of dead specimens

When an animal has died, it is important to provide for the optimal use of the materials available, particularly for studies, which may benefit the species. Tissue collection and post-mortem protocols, and addresses of recommended recipients of material are available from the BIAZA Office or relevant Taxon Working Group and EAZA TAG chairs.

F. Veterinary screening

The BIAZA guidelines on minimising the risk of disease transfer between member collections should be followed (Appendix 5). However the Balai Directive (Council Directive 92/65 (BALAI)-(amended Council regulation 1282/2002)) will affect the requirements for veterinary screening when moving animals between approved institutions in the EU.

III. Use of Animal Dealers in Animal Transactions

Animal dealers/brokers should be used only when:

- (a) the name and address of the potential new owner is provided by the broker/dealer;
- (b) the potential new owner states in writing interest in the animal;
- (c) before any animal is transferred, the potential new owner is approved by the sending zoo or another BIAZA member, or the BIAZA or the EAZA Offices *or an alternative suitable reference*, and information given about the enclosure in which the animal will be housed demonstrates that the facility is suitable;
- (d) the sending institution handles all eventual CITES documentation and identification;
- (e) the transportation and the transport crates are arranged in discussion with the broker/dealer and **MUST** comply with relevant legislation, such as the Welfare of Animals (Transport) Order 1997, or the regulations of the International Air Transport Association (IATA);
- (f) the new owner or the dealer informs the promptly of the arrival of the correct animal.
- (g) in the rare cases when animals are acquired from dealers, full details of their source and means of acquisition **MUST** be obtained.

Appendix 1. Minimising Surplus

Husbandry techniques considered appropriate:

N.B. Not all techniques may be appropriate for all species, and should be considered on a case by case, species by species basis.

- only hand-rearing for pre-determined population management purposes (e.g. according to TAG or EEP recommendations);
- separation of males and females
- holding single-sex groups;
- extending interval between births;
- removing, shaking or freezing eggs;
- euthanasia (including culling).

Veterinary techniques considered appropriate:

N.B. Not all techniques may be appropriate for all species, and should be considered on a case by case, species by species basis.

- Contraception may be used wherever there is a need for reasons of population management. However, the possible side effects of both surgical and chemical contraception, as well as the negative impact on behaviour, should be considered before a final decision is made.
- hormone injections or implants in females;
- hormones given orally to females;
- PZP vaccinations of females (not yet licensed in the UK);
- sterilisation of males/females, including vasectomy and castration;
- abortion.



Appendix 2 *Pro forma - Animal Transfers*

When a Member is sending an animal(s) to another collection or individual that is not known to them, the sending institution **MUST** ensure that they have either inspected the facilities at, or received suitable references concerning, the receiving collection. Such reassurances, regarding facilities and expertise, are **MANDATORY** for all species included in Category I of the *Dangerous Animal Categorisation* under the *Zoo Licensing Act* and the *Secretary of State's Standards of Modern Zoo Practice (SSSMZP)*, all primate species, and other species that Council may from time to time nominate. The use of the following form alone is not considered adequate reassurance for the transfer of these animal groups.

Whilst this high level of scrutiny should be applied for all taxa if possible, it is recognised that for certain taxa a lower level of reassurance may be appropriate. In these cases Members should use this approved form to establish a minimum level of information prior to implementing the transaction.

This form must also be used when any species is being sent to a zoo, aquarium, institution or individual that is NOT a member of BIAZA or a member of a recognised regional zoological association (e.g. EAZA, AZA etc.)

The intended recipient **MUST** complete and return the form to the BIAZA Member **BEFORE** any transaction can take place as described in the above circumstances. Similarly, if an intermediary is used to transfer the animal(s), they must have the form completed by the recipient(s) of the animal(s) and return the completed forms to the BIAZA Member.

Please note: This form MUST be kept on file for the duration of the standard Zoo Licence (UK Zoo Licensing Act (1981)), i.e. for a minimum of six years.

PART ONE – To be completed by sending institution

Name of BIAZA member collection (Sender):	
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Species to be transferred	
Scientific Name:	
Common Name:	

If more than one species is being transferred in the transaction, **a separate form will need to be filled out for each species.**

Number of individuals to be transferred (include ARKS ID#s)		
Male	Female	Unsexed

Has all necessary paperwork been checked for this transaction? (e.g. Article 10, Article 60, CITES, DWA) Give details.	
---	--

Have these specimen(s) been posted on the BIAZA and/or EAZA Available List? Give details.	
--	--

Are these specimen(s) part of a managed programme? If so, give details.	
--	--

If the specimen(s) are part of a managed programme, has the necessary permission been obtained for this move from the programme coordinator?	
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What are the terms of this transaction i.e. donation, loan, sale, exchange? Give details.	
--	--

PART TWO – To be completed by intended recipient

Name of Recipient:	
Business Name (if applicable):	
Address:	
Telephone:	
Fax:	
Email:	

Is this the final destination of the specimen(s)? If, 'NO', then please give full details here.	
--	--

Recipient's Veterinary Surgeon:	
Veterinary Practice:	
Address:	
Telephone:	
Fax:	
Email:	

Details of recipient's previous experience with this species or similar species.	
---	--

Full details of accommodation <i>(include indoor, outdoor dimensions, heating, lighting, substrate, furniture etc.)</i> Also include sketches or photographs of intended accommodation.	
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Method of transportation <i>(include how long the animals are likely to be in transit).</i>	
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Proposed date of transfer	
----------------------------------	--

PART THREE - Declaration

I confirm that the above information is, to the best of my knowledge, complete and correct.

For sending collection		For recipient	
Signed:		Signed:	
Full Name:		Full Name:	
Position:		Position:	
Date:		Date:	

Appendix 3.

Useful References

ANIMAL HEALTH AND WELFRARE (Scotland) ACT 2006. <http://www.scotland.gov.uk/Topics/Agriculture/animal-welfare/AnimalWelfare/AHWSAGuidance/AHWSAGuideintro>

ANIMAL WELFARE ACT 2006 (England and Wales) <http://www.defra.gov.uk/animalh/welfare/act/info.htm>

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IUCN (2000b) Guidelines for the Placement of Confiscated Animals. IUCN/SSC Reintroduction Specialist Group. <http://www.iucn.org/themes/ssc/sgs/rsg/rsgcdrom/PDFs/EnglishConfGlines.pdf>

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Appendix 4. EAZA/EEP CONDITIONS OF ANIMAL TRANSFERS AND THE ROLE OF THE SPECIES COORDINATOR IN TRANSFERS

Conditions of transfers

Participants will not transfer an EEP animal without prior approval of the Species Coordinator

Transfers of EEP animals between EEP participants can be performed under one of the following conditions:

1. Donation (an animal X is made available – free of charge – by participant A to participant B, who becomes the new proprietor).
2. Exchange (animals X and Y are exchanged between participants A and B, who become the new proprietors of the newly received animals; X and Y do not necessarily belong to the same [EEP] species).
3. Loan (animal X is transferred from participant A to B, but A remains its proprietor; agreements can be made as to the ownership of the offspring of X).

It is recommended that participants involved in a transfer choose from options 1-3. In exceptional cases option 4 can be chosen:

4. Sale (animal X is sold by participant A to participant B, who becomes the new proprietor)

It is strongly recommended, however, to choose conditions that do not put up such financial barriers that transfers recommended for population management may become impossible.

[N.B. in May 2003 EAZA Council approved the following: *‘For the benefit of the future viability of EAZA/EEP populations, all transfers of EEP animals must be arranged in full consultation with, and the agreement of, the EEP Co-ordinator. In order to ensure the non-commercial status of EEPs any selling of EEP animals must be avoided’.*]

Transfers between participants and non-participants

Participants will not transfer an EEP animal to a non-participant without prior approval of the Species Coordinator. Approval will be given if the animal concerned is surplus to the EEP population. If the animal is not surplus to the population it can only be transferred to the non-participant if the latter: 1. is resident in the EEP region, and 2. is willing to join the EEP programme for the species and to abide by its rules.

Transfers of animals from non-participants to participants also need approval by the Species Coordinator, who will grant permission only if such animals are considered valuable to the EEP population.

Transfers recommended for population management

Transfers of animals for management of the EEP population will be recommended and by the Species Coordinator, and approved by the Species Committee on an annual basis (unless species-specific factors require more frequent, or warrant less frequent recommendations). The Species Coordinator contact all participants involved in these transfers and stimulates their timely implementation.

Transfers suggested by participants

Participants may suggest additional transfers, not specifically recommended for population management. In such cases they will always contact the Species Coordinator, who will study the effects of these transfers on population structure. The Coordinator will grant permission if there are no negative effects; alternative transfers will be proposed if negative effects are expected.

Placement of surplus

Transfers should not be arranged via brokers or dealers. Participants should directly contact each other, and if necessary the Species Coordinator to suggest the best possible solution in the light of proper population management. Only if the Coordinator does not have a clear suggestion, possible recipients of surplus animals may be sought by placing them on circulated surplus lists of the participants, and/or on the 'available/wanted' list published regularly by EAZA. A code should then be added indicating that transfers need approval of the EEP coordinator.

The role of the species coordinator

Apart from the role of the Species Coordinator in animal transfers as indicated in the above paragraphs, the Coordinator may act as an intermediary between two participants in the implementation of recommended transfers. Strictly speaking the agreement on the conditions of a recommended transfer is a matter for the participants involved. However, if the latter do not arrive at an agreement on the terms of an important transfer, the Coordinator may try to bring the parties together, or – if this turns out to be impossible – the Coordinator will try to find the next best solution for population management by involving a third or a fourth party in the transfer. If no acceptable alternative is found the Coordinator may put the matter to the Species Committee to make a decision.

In cases of transfers between an EEP participant and a non-participant outside of the EEP region the Species Coordinator will contact the Species Coordinator of the region of the non-EEP participant if there is a formal breeding programme for the species in that region. The coordinator will then make sure that the suggested transfer will not interfere with the management plan in that region.

Appendix 5

. BIAZA Guidelines on Minimising the Risk of Disease Transfer between Member Collections

Introduction:

The purpose of these guidelines is to set down general principles of veterinary surveillance to which a collection worthy of being a BIAZA member should aspire.

With this in mind a sending institution has a duty of care to ensure that any animals transferred are, as far as can reasonably be ascertained, healthy and fit for purpose. No animal showing clinical signs of disease should be moved between collections unless the condition in question is chronic in nature and the receiving collection is willing and able to continue to manage the animal in appropriate facilities.

Diseases of concern are likely to change with time such that it is the intention that these guidelines and the Appendix be reviewed annually.

General Principles:

1. Any animal move carries with it a risk of disease transfer.
2. These diseases may be infectious or non-infectious.
3. **Infectious diseases** may cause problems in the individuals being transferred, their conspecifics, other species in the collection or in humans (staff and/or visitors).
4. **Non-infectious diseases** (including behavioural abnormalities) tend to affect only the health and welfare of specimens being transferred but they may also have other knock on effects (e.g. suitability for breeding if the animal is infertile due to testicular abnormality, suitability for enclosure type available if the individual cannot move normally etc).
5. The aim of this document is to provide guidance as to how to minimise the risk of disease transfer between BIAZA collections. For the majority of moves this will be very straightforward.
6. The most important techniques for minimising disease transfer are:
 - a. Pre-export health screening
 - b. Quarantine and post-import health screening
7. Both of these techniques should be seen as routine for all animal moves, but this document will focus on disease screening (quarantine protocols are covered elsewhere).
8. Effective disease screening generally requires one to know what one is looking for and what the significance of finding it is.
9. Some tests (such as physical examination or haematology/biochemistry) will be broad spectrum and can pick up a range of different abnormalities, but most tests are very specific hence the list of diseases of concern needs to be decided first.
10. The diseases of concern may vary from one move to the other as they are dependent on many factors including:
 - The species being transferred.
 - The disease history and adequacy of the veterinary surveillance programme of the sending collection.
 - The disease history of the receiving collection.

- The purpose of the animal in the receiving collections (e.g. for handling sessions with the public, for breeding, as part of a mixed species exhibit).
- The current UK/regional disease status
- The suitability of post import quarantine facilities

The following sections outline:

Section A: BIAZA recommendations for pre-transfer disease screening: minimum standards

Section B: Disease Risk Analysis: guidelines on the risk assessment process

Appendix: Potential Infectious Diseases of Concern for Transfers within the BIAZA Region: arranged taxonomically and including justification as to why these diseases (excluding those which are non-infectious) should be considered and how they might be screened for.

A. BIAZA Recommendations

- As a minimum before sending an animal to another institution all members should:
 - Submit a full medical history of the animal to be transferred AT LEAST 1 WEEK PRIOR TO TRANSFER to the receiving collection. In the absence of a medical history, as a minimum a written declaration stating that the animal being transferred appears to be in good health and that there have been no known recent problems with it or its conspecifics should be sent to the receiving collection.
 - Notify the receiving collection AT LEAST 1 WEEK PRIOR TO TRANSFER of any disease concerns in its immediate group / or in the collection as a whole.
 - Where practicable carry out a physical examination of the animal within 7 days of transport (by a vet) and a visual examination by a vet and/or experienced person with the species in question within 24 hours of transport/upon departure.
 - Faecal parasitology and bacteriology (depending on medical history)
- Additional disease screening for certain taxa is recommended (see Appendix).
- Screening requirements should be agreed between the sending and receiving collections.
- Test availability and impact on the animal to be transported must also be considered. A good health history, including details of any new imports to the group and results of post-mortem examinations over a period of years, may prove adequate.
- Liability for screening costs to be agreed between the parties involved.
- Pre-export screening does not replace the need for post-import quarantine. As a rule of thumb mammals, birds and fish should be isolated from the rest of the collection (or co-terminously with other conspecifics if preferable for social taxa) for a minimum of 30 days; 90 days is recommended for reptiles.

B. Disease Risk Analysis

Disease risk analysis should be performed by the receiving institution's vet in partnership with the animal management staff. Though the process may seem involved the first time round, many transfers are very similar in make up and a pre-export testing schedule for many of a collection's routine imports from their key partners should quickly emerge. These tables need only then be worked through for more unusual ones.

Questions	Considerations
Q1. What groups might be at risk?	<ul style="list-style-type: none"> • animal being transferred • animals of the same species already in the collection • animals of different species which may come in contact with the imported animal either directly or indirectly

	<ul style="list-style-type: none"> humans (staff and/or visitors)
Q2. What are the infectious disease agents that this species might be harbouring?	<ul style="list-style-type: none"> See Appendix for some of the more important diseases of this species. Also consider diseases of in-contact species that this individual might also be carrying (e.g. mechanical transfer of chytrid fungus between collections)
Q3. What non-infectious health issues might the individuals to be imported be harbouring?	<ul style="list-style-type: none"> Examples might include chronic foot problems, teeth problems, poor fertility, metabolic bone disease, drug or food intolerances, heart disease Behavioural health should also be considered (e.g. History of infanticide, abnormal levels of aggression to other animals or staff).
Q4. What is the likelihood that the animal to be transferred is harbouring these diseases / disease agents?	<ul style="list-style-type: none"> Consider current diseases of concern in the UK or region. (e.g. TB, avian influenza, tetanus etc). Closed collections (i.e. ones without any recent imports to the group) are much less likely to be incubating infectious diseases Measures to decrease the likelihood of disease transfer include pre-export prophylactic treatment (e.g. worming, vaccination).
Q5. What is the potential significance of each of these diseases / disease agents to each of the risk groups?	<ul style="list-style-type: none"> See Appendix for guidance. The significance of some disease issues might be decreased by adjusting the management practices at the receiving collection (e.g. handling chutes to allow training for conscious foot care, avoidance of certain drugs that the animal has reacted badly to). Be aware that some pathogens might not cause disease in your collection but, if they are detected, they might lead to restrictions on animal moves (e.g. presence of low pathogenic strains of avian influenza may shut down the zoo)
Q6. Can the diseases / disease agent that are both significant and likely, be screened for before export?	<ul style="list-style-type: none"> Not all diseases can be screened for: The diagnostic test may not have been developed, may not be routinely available, may not be accurate, may require samples that are difficult or dangerous to obtain or may be prohibitively expensive. The Appendix provides guidance as to whether a diagnostic test is available and what samples might be required.
Q7. If they can't be effectively screened for, are there any other measures that could be taken to reduce the risk?	<ul style="list-style-type: none"> For those diseases where no diagnostic tests are practicable a combination of medical history (including PM's) and post import isolation may be the best protection. Prophylactic treatment may be useful in some instances (see Q4)

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APPENDIX: Potential Infectious Diseases of Concern for Transfers within the BIAZA (British & Irish) Region

MAMMALS

ALL ANIMALS SHOULD HAVE:

- **Medical history sent a minimum one week prior to export**
- **Declaration of presence or absence of declaration diseases**
- **Prophylactic treatments as recommended**
- **A physical examination – including notification of findings to receiving collection**

IT IS HIGHLY RECOMMENDED THAT ALL ANIMALS SHOULD HAVE:

- **Tests for the diseases of concern indicated**

Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY-HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
Primates	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> • Tuberculosis (bovis or tuberculosis) in previous 5 YEARS • Suspicious reactors to TB skin test in previous 12 months. • Animals testing positive for Herpes B (macaques) • Animals testing positive for Hepatitis B 							
	Enteric nematodes (highlighting Strongyloides, pinworm)	Zoonoses. Known to cause morbidity in NHP's	M	M	N Treat	Faecal parasitology -3 day pooled sample. Strongyloides may require charcoal culture for ID	Faeces	3 day pooled sample (plus history of this being done). cancan be difficult to pick up (intermittent shedding) so treatment should be considered prior to a move even if test negative
	Enteric protozoa (highlighting E.histolytica, B.coli, B.hominis, D.fragilis)	Most zoonoses. Confirmed clinical disease and carrier states in majority of NHP. Severe under reporting of protozoal infections in UK zoos suspected	M	H	N	Fresh stained faecal smear. Fresh-frozen faeces for E.histolytica	Faeces	Sample twice, 1 week apart.. Samples must be very fresh.. If no in house ability, can put faeces in formalin for lab analysis of any cysts.
	Enteric bacteria (Highlighting Shigella (apes),	Zoonoses – known to cause morbidity and occasionally mortality in NHP's	M/H	M	Y potentially	Bacteriology	Faeces	Salmonella should be typed and Campylobacter speciated, especially in subclinical carriers

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY-HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
	Salmonella, Campylobacter)							
	Tuberculosis caused by M. bovis or tuberculosis	Important cause of mortality/morbidity and ZOOONOSIS	H	L	Y	Y – skin test, also gamma interferon blood test for some species Culture gold standard but slow and insensitive	First line standard: Skin test May also consider: Tracheal/ bronchial wash for culture Serum/ plasma (Investigating TB antibody Stat-Paks from Chembio)	Highly recommended though might be acceptable to forgo if closed group with regular negative testing. NEED TO DISCUSS REGIME WITH RECEIVING COLLECTION Remember to consider TB status of in contact humans. If animals are in a walkthrough exhibit or there is any chance their carers may have been infected, testing is highly recommended.
	Hepatitis B	potential zoonosis	L	M	N but vacc. keepers	Y - Virus Isolation/ PCR/ ELISA etc	Blood (Serum)	Gibbons common carriers. See TAG for notes on management.
	Herpes Viruses eg: simplex (apes) ateles (spider and owl monkeys) B (macaques)	Herpes B potential Zoonosis. Other herpes viruses can cause fatal disease in aberrant primate species)	H/L (sp. Dependan t)	M	Herpes B Y Others N but sp dep.	Y - Virus Isolation/ PCR/ ELISA etc	Blood (Serum)	HPA reference laboratory for Herpes B. – can also screen for presence of other alpha herpes viruses but may not be able to identify to species level.
Ruminants and camelids	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths) <ul style="list-style-type: none"> • Tuberculosis/Mycobacteriosis. (in previous 3yrs) • Paratuberculosis (Johnes Disease - in previous 2yrs) • Transmissible spongiform encephalopathies (in previous 10yrs). • Lumpy Jaw • Foot rot • Bluetongue • Domestic Cattle on premises: History of BVD/MD, IBR, Leptospirosis. • Trichuris (especially for camels) 							
	Endoparasites. Helminths, protozoa etc	Important cause of disease/morbidity	M/H	H	N TREAT	Faecal examination. Pooled faeces from group is probably acceptable although individual samples better	faeces	Faeces collected over several days better than an individual sample

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY-HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
	Salmonellosis/ Campylobacter	Important cause of disease/morbidity and ZOOONOSIS	M	M	N	Faecal cultures. Pooled faeces from group is probably acceptable	faeces	Faeces collected over several days better than an individual sample
	Tuberculosis	Important cause of disease/morbidity and ZOOONOSIS.	H	L	Y	Intradermal skin test Possible gamma interferon blood test available		Test could be omitted on basis of collection history and local disease status. To be discussed between vets for both collections
	Mycobacterium paratuberculosis (Jones Disease)	Important cause of disease/morbidity and ZOOONOSIS.	H	L	Y	ELISA and AGID available for cattle. CFT and AGIDT available for sheep and goats. Faecal exam unreliable	Blood	Appropriate test for species to be discussed between vets and VLA. Vaccination may affect tests. Faecal culture is gold standard to eliminate atypical mycobacterial infection
	Malignant Catarrhal fever	Important cause of disease in deer, antelope and cattle. Sheep can be symptomless carriers	H	M	Y	IFAT and SNT available.	Blood	sheep, wildebeest and other alcelaphine antelope should be screened if planning mixing or accommodating near sensitive species such as Pere David Deer
	MV/CAE	Important cause of disease in sheep and goats.	H	M	Y	ELISA	Blood	Sheep and goats only
Pigs Peccaries and hippos	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> • Tuberculosis/mycobacteriosis (previous 3yrs) • Domestic pigs on premises: PRRS, Atrophic Rhinitis, Parvovirus, Mycoplasma hyopneumoniae, PMWS/PDNS, TGE, Strep suis meningitis.. 							
	Endoparasites. Helminths, protozoa etc	Important cause of disease/morbidity	M-H	H	N TREAT	Faecal examination. Pooled faeces from group is probably acceptable although individual samples better	faeces	Faeces collected over several days better than an individual sample
	Salmonellosis/Campylobacter	Important cause of disease/morbidity and ZOOONOSIS	M	M	N	Faecal cultures. Pooled faeces from group is probably acceptable	faeces	Faeces collected over several days better than an individual sample

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY-HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
	Tuberculosis	Important cause of disease/morbidity and ZONOSIS in Hippos Questionable justification in pigs and peccaries at this time	H	L	Y	Intradermal skin test not reliable/validate. Possible blood test available	Blood	Testing should be discussed with receiving collections and with TAG. May be recommended in Hippos depending on history of population / individual.
Equidae	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> • Contact with domestic horses: influenza, strangles, CEM • EHV, EIA and sarcoid • Must have a Horse Passport (mandatory from July 09) 							
	Enteric parasites. Strongyles in particular and strongyloides, parascaris, oxyuris, spirurids, tapeworms and cyathastomes	Common, can be important cause of morbidity	M	H	N TREAT	Various methods of quantitative and qualitative faecal tests and hatching of eggs into larvae with subsequent identification	Quantitative egg count on 3 consecutive day faecal sample. Need 3-5g faeces	Faeces collected over several days better than an individual sample Where risk of occult cyathostome infections, special treatment regimes are needed
	Salmonella spp.	Zoonotic disease	M	L	N	Culture (+/- serotyping also PCR)	Faecal sample on 3-5 consecutive days. Need 3-5g faeces.	Faeces collected over several days better than an individual sample
Tapir and Rhino	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> • Rhino: skin disease • Tuberculosis 							
	enteric parasites	common in white rhino with little clinical disease	M-L	M-H	N	Faecal examination.	faeces	Pooled faeces from group is probably acceptable although individual samples better
	Tuberculosis	Important cause of disease/morbidity and ZONOSIS	H	L	Y	Refer to TAG recommendations		TB infection is a reported problem in Tapirs. Definitely worth considering.
	faecal bacteriology		M	L	N	Faecal cultures.	faeces	Faeces collected over several days better than an individual sample Pooled faeces from group is probably acceptable
Elephants	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> • EEHV 							

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY-HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
		<ul style="list-style-type: none"> Tuberculosis / mycobacteriosis (previous 3yrs) Elephant pox 						
	Endo-parasites	Cause of morbidity	L	M-L	N Treat	Faecal parasitology	Faeces (3 samples over 3 weeks)	
	Salmonella	Cause of mortality/morbidity and ZONOSIS	M	L	N	Faecal culture	Faeces (3-5 consecutive days worth of samples cultured separately)	Carrier animals may only shed intermittently Faeces collected over several days better than an individual sample
	TB (M.tuberculosis or M. bovis)	Important cause of mortality/morbidity and ZONOSIS Treatment options very limited	H	L	Y	Trunk washes for culture still definitive test despite 8 week wait and lack of sensitivity Rapid Test (RT) and confirmatory Multiprint Immunoassay (MAPIA) technology appear to show upto 100% sensitivity and much earlier diagnosis	Multiple trunk wash samples (at least 3 within 7 days) Whole blood, serum or plasma	Intra-dermal skin test demonstrates very poor sensitivity RT and MAPAI will replace culture once validity further demonstrated Refer to elephant TAG for current testing recommendations
	EEHV (Elephant Endotheliotropic Herpesvirus)	Many viruses circulating. No disease in most individuals BUT can be Important cause of mortality/morbidity with peracute course in naïve individuals no vaccines or well proven therapy	H-L	H?	??N	Serology possible in Europe or USA but needs co-ordination by someone! PCR on blood of clinical cases or PM tissues of other herd members	Plasma (preferred) or serum (can be frozen and sent as batches)	Current thinking suggests that all elephants are infected with one or more strains. Introduction of a new strain may cause peracute disease in naïve individuals. Refer to Elephant TAG recommendations (currently under review)
Rodents, insectivores and lagomorphs and sloths	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> Sendai virus Sialodacryoadenitis EMCV (encephalomyocarditis virus) 							

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY-HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
	<ul style="list-style-type: none"> LCMV Yersiniosis Capillaria hepatica <p>Recommended pre-export prophylactic treatments:</p> <ul style="list-style-type: none"> Vaccination of Lagomorphs against myxomatosis and viral haemorrhagic disease 							
	Enteric parasites (and fascioliasis in beavers)	Common,	M	H	N Treat	faecal smears , McMasters, identification of cysts or trophozoites in smears, serology for giardia	3 consecutive days faecal sample	Faeces collected over several days better than an individual sample Pooled faeces from group is probably acceptable although individual samples better
	Enteric bacteria (eg salmonella)	causes disease AND zoonotic	M	M	N	Y	Faeces / cloacal swab	Highly recommended
	Encephalitozoon cuniculi (lagomorphs)	Endemic in captive population of lagomorphs; infection of rodents possible	M	H	Y treat	serology	blood	Testing to determine positive or negative status in lagomorphs recommended; treatment available
	Sarcoptic mange, lice and other ectoparasites	Can be debilitating leading to morbidity and mortality and contagious	M	M	Y treat	microscopy /hair samples	Skin scraping / hair pluck/ tape strip / physical exam	All animals with significant ectoparasite burdens should be checked for other underlying disease
	Mycoplasma (rats)	important cause of respiratory disease	M	M	N	Y	nasal swab	
	Lymphocytic choriomeningitis LCMV (small rodents)	Zoonoses, can be spread to callitrichids and easily transferred from wild rodents to captive rodents	H	M	?Y depends on status of collection	serology	Blood/serum from individual or from small proportion of the group.	
Chiroptera	<p>In-hand health-check to confirm ID and check general health including teeth, patagial integrity, limbs and external genitalia. Radiography to be performed if legs or wings give concern.</p> <p>Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)</p> <ul style="list-style-type: none"> Lyssavirus (in previous 3yrs) – include screening history ie number of samples submitted over the previous 3yr period – this is particularly important for any walk through exhibits. 							
	Intestinal parasites	Important cause of morbidity and mortality	L	L	N treat	Routine faecal flotation	Faeces	Three samples at weekly intervals from known individuals if small

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY-HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
								group or pooled sample from large group
	External parasites	Important cause of morbidity	L	L	N treat	Visual check/sticky tape and/or skin scrape	Skin/hair or skin debris	Single sample from unaffected animals at health check
	Faecal bacteria	Important cause of morbidity and mortality	L	L	N but depends if walk through...	Microbiology	Faeces	Three samples at weekly intervals from known individuals if small group or pooled sample from large group – ensure screened for zoonoses including <i>Salmonella</i> and <i>Campylobacter</i>
	Lyssavirus	Important cause of mortality/morbidity and zoonosis	H	L	Y	FAVN test /	Serum / also submit heads of any bat that dies to VLA for screening	Test performed at VLA Weybridge. Would recommend test on all if a small group or if destined for a walk-through exhibit, otherwise a representative sample
Marsupials	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> Lumpy Jaw 							
	Enteric parasites	Coccidiosis common in joeys. Monitor for nematodes	M	M	N TREAT	Faecal parasitology	Faeces	3 day pooled sample
	Salmonella and campylobacter	Zoonosis and can cause severe morbidity	M	M	N	Bacteriology	Faeces	Type Salmonella whenever possible
	Lumpy Jaw (Bacteroides/ Fusobacterium)	Common cause of morbidity	M	M	N	Clinical signs? Bacteriology	Lesion swab	Test only when suspect case
	Chlamydia	Common in Koalas. Zoonosis.	L	L	N	'Clearview' rapid test	Blood	Koalas only
Felidaea	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> Any of the diseases listed below 							
	Recommended pre-export prophylactic treatments:							
	<ul style="list-style-type: none"> Vaccinations for FELV, FIV,+ other feline viruses up to date (full vaccination history and opportunistic testing may negate need for testing prior to export – discuss with receiving collection). NOTE Beware use of modified live vaccines in non-domestic felids. 							
	enteric parasites		M-L	M-H	N	Faecal examination.	faeces	Pooled faeces from group is probably acceptable although individual

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY-HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
								samples better
	faecal bacteriology		M	L	N	Faecal cultures.	faeces	Faeces collected over several days better than an individual sample Pooled faeces from group is probably acceptable
	Feline Immunodeficiency Virus (FIV)	Potential cause of serious immunodeficiency-like disease	H/M	L (except lions)	Y	Antibody testing by ELISA & Western Blotting	Serum	Possibly prolonged seroconversion times in non-domestic species.
	Feline Leukaemia Virus (FeLV)	Potential cause of neoplastic & degenerative conditions	M	L	Y	Antigen test	Serum	Domestic cat vaccines not validated in non-domestic species
	Feline Coronavirus	Potential cause of fatal Feline Infectious Peritonitis	M	L	N	Antibody test. (PCR for virus shedding currently unavailable in the UK)	Serum	Interpretation of antibody titres complicated. Seek veterinary advice if positive
	Chlamydomphila felis	Cause of ocular & respiratory disease, and possibly involved in infertility	M	M	N	PCR	Conjunctival swab	Vaccination probably effective
	Feline Herpes Virus (FHV)	Cause of severe respiratory disease, ulcerative keratitis & dermatitis	H (if not vac)	M	Y	PCR & virus isolation	Oropharyngeal swab in VTM	Carrier status recognised. Vaccination effective
	Feline Calicivirus (FCV)	Cause of severe oral & respiratory disease (and lameness)	M	M	Y	PCR & virus isolation	Oropharyngeal swab in VTM	Carrier status recognised. Vaccination variably effective due to rapid virus evolution.
Canidae	<p>Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)</p> <ul style="list-style-type: none"> Distemper, parvovirus, adenovirus, leptospirosis, sarcoptic mange <p>Recommended pre-export prophylactic treatments:</p> <ul style="list-style-type: none"> Vaccinations up to date for canine distemper, parvovirus, adenovirus 1 and leptospirosis 							
	Endoparasites Nematodes Cestodes Coccidia (Neospora caninum)	Common, some zoonotic	M	H	N	Faecal flotation, including Baermann technique for lungworm	Faeces	Test pre-move and treat with appropriate anthelmintics/anticoccidials pre-move. If positive inform receiving zoo, which should also test on arrival and re-treat during quarantine period

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY-HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
	Ectoparasites Fleas Mites/Lice/Ticks	Can cause morbidity. Can be involved in transfer of infectious agents e.g ticks and <i>Borrelia</i>	L	M	N	Visual examination Skin scrape	skin scrape. Visual exam	Treatment indicated with appropriate ectoparasiticide if ectoparasites detected on pre-move physical examination. Any animals with skin lesions should be investigated pre-move
	Salmonella	Common, can cause morbidity. Zoonosis	M	M	Variable	Culture	Faeces	Test pre-move, only if animal has abnormal faeces. If positive, discuss significance with receiving zoo's vet. NB Healthy animals with normal faeces highly unlikely to be positive. Treatment with antibiotics generally only indicated if risk of contact with immunosuppressed people/animals or children, and may induce latent carrier status
Mustelids/ viveridae/ procyonidae	<p>Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)</p> <ul style="list-style-type: none"> • distemper <p>Recommended pre-export prophylactic treatments:</p> <ul style="list-style-type: none"> • Consider use vaccines against canine disease but beware use of modified live vaccines as may cause disease in these species. 							
	enteric parasites		M-L	M-H	N	Faecal examination.	faeces	Pooled faeces from group is probably acceptable although individual samples better
	faecal bacteriology		M	L	N	Faecal cultures.	faeces	Faeces collected over several days better than an individual sample Pooled faeces from group is probably acceptable
Marine mammals	<p>Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)</p> <ul style="list-style-type: none"> • Tuberculosis • Morbillivirus • Herpes virus in seals • Small pox • Brucellosis 							
	enteric parasites incl. lungworm		M-L	M-H	N	Faecal examination.	faeces	Pooled faeces from group is probably acceptable although individual

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY-HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
								samples better
	faecal bacteriology		M	L	N	Faecal cultures.	faeces	Faeces collected over several days better than an individual sample Pooled faeces from group is probably acceptable
	morbillivirus		H	L	Y	Serological and/or PCR	blood	
	herpes (seals)		H	M	Y? possible	Serological and/or PCR	blood	Data deficient in most but does affect common seals seriously
	Brucella		Zoonotic hazard	M	N?	Serology at VLA	blood	
	Tuberculosis		H	Species related	Y	Skin though not reliable. New rapid tests		Particularly some sealions and South American fur seals

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BIRDS

ALL ANIMALS SHOULD HAVE:

- Medical history sent a minimum one week prior to export
- Declaration of presence or absence of declaration diseases
- Prophylactic treatments as recommended
- A physical examination – including notification of findings to receiving collection

IT IS HIGHLY RECOMMENDED THAT ALL ANIMALS SHOULD HAVE:

- Tests for the diseases of concern indicated

Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
Passerines	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths) <ul style="list-style-type: none"> • Chlamyophilosis • Atoxoplasmosis 							
	Enteric nematodes	Big cause of morbidity and occasionally mortality	M	H	N TREAT	Parasitology	Faeces	3 day pooled faecal sample
	Atoxoplasma	Known infection in the UK. Several species – causes high mortality – fledglings usually, but Bali mynah particularly sensitive	H	M/H	Possibly Y (dependant on sp.)	Faecal parasitology for oocyst detection (multiples required)	Faeces +/- blood buffy coat	Will be species dependant. At least 3 negative faecal samples required at one week intervals if using parasitology alone.
	Salmonella and Campylobacter	Zoonosis	M	?M	N	Bacteriology	Faeces	Salmonella positives should be typed
	Chlamyphilila	Zoonosis. Found in UK collections.. Can cause debilitation.	M	M/L	Y TREAT	PCR	Heperanised blood, Faeces or cloacal swab	Only if history at collection within the previous 12 months. Single sample required.
	Avian Polyoma virus (Gouldian finches)	Species specific?	H	M	?Y	PCR	Feather, heperanised blood, faeces, cloacal swab	Single sample required
Falconiformes	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths) <ul style="list-style-type: none"> • Avipox: Spread by flies. Declare if cases in previous 12mths, suggestive clinical signs or if the bird has been imported or has been housed with/near birds imported from Middle East.. DX by EM/histopath of lesions. Serology also possible. • Chlamyophilosis 							

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
<p>Recommended pre-export prophylactic treatments:</p> <ul style="list-style-type: none"> Aspergillosis: Important cause of morbidity in stressed raptors especially gyrs (and their crosses), goshawks, Snowy Owls and mountain eagles (eg Golden Eagles) Use of itraconazole at 10mg/kg sid po recommended in all susceptible species for 7-10 prior to move until 2 weeks post-moveNote:, if the administration of drug will cause more stress or if the bird is paired with another (thus making administration unreliable) then it may be wise to ignore this 								
	Endoparasites nematodes -coccidia	Coccidia esp in falcons, esp merlins and their hybrids	M	H	N Treat	Faecal floatation	Faeces	Would recommend at least one sample pre- and post- move – the latter being 7-14 days after move during quarantine period Faecal samples should be pooled 3-day samples for coccidia
	Chlamydophilosis	Many wild raptors appear to be seropositive Therefore worth considering in passage birds or in those that have had exposure to wild birds – eg used for hunting	M	L/M	Y Treat	Serology to assess exposure Faecal PCR to assess shedding	Blood / Faeces	Serological test – if positive then perform PCR
Waterfowl	<p>Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)</p> <ul style="list-style-type: none"> Avian TB Yersiniosis Chlamydophilosis 							
	Enteric Parasites	major cause of debility	M/H	H	Treat	Y	Faeces	Highly recommended
	Enteric bacteria (eg salmonella)	causes disease AND zoonotic	L	H	N	Y	Faeces / cloacal swab	Highly recommended
Psittacines	<p>Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)</p> <ul style="list-style-type: none"> Chlamydophilosis PBFD Polyoma Psittacine Herpes Virus (Pacheco's Disease) Proventricular Dilatation Disease 							
	Enteric nematodes	Big cause of morbidity and occasionally mortality	M	H	TREAT	Parasitology	Faeces	3 day pooled faecal sample

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
	Salmonella and Campylobacter	Zoonosis	M	?M	N	Bacteriology	Faeces	Salmonella positives should be typed
	Chlamydomphila	Zoonosis. Can cause debilitation.	M	M/L	Y TREAT	PCR	Heperanised blood, Faeces or cloacal swab	Highly recommended.
	Psittacine Beak and Feather Disease (Pbfd)		H	M	Y	PCR	Heperanised blood, Feather pulp	Highly recommended.
	Polyoma Virus		H	M	Y if receiving collection is free	PCR	Heperanised blood, Faeces or cloacal swab	Highly recommended. Lovebirds can be latent carriers.
	Psittacine Herpes Virus		H	M	Y	Serology (VLA) but not very sensitive. Autopsy only reliable test	Blood	Species specific
Columbiformes	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> Chlamydomphila. 							
	Enteric nematodes	Big cause of morbidity and occasionally mortality	M	H	N TREAT	Parasitology	Faeces	3 day pooled faecal sample
	Trichomonas/ Candida	Known morbidity in the UK	M	M	N Treat	Crop swab – examine warm on microscopy	Crop swab	Only if suspect on clinical examination
	Salmonella and Campylobacter	Zoonosis	M	?M	N	Bacteriology	Faeces	Salmonella positives should be typed
	Chlamydomphila	Zoonosis. Found in UK collections.. Can cause debilitation.	M	M/L	Y TREAT	PCR (BioBest)	Heperanised blood, Faeces or cloacal swab	Only if history at collection within the previous 12 months. Single sample required.
Penguins	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> Plasmodium (Pododermatitis) 							
	Recommended pre-export prophylactic treatments:							
	<ul style="list-style-type: none"> Aspergillosis: Important cause of morbidity in stressed penguins. Use of itraconazole at 10mg/kg sid po recommended in all susceptible species for 7-10 prior to move until 2 weeks post-move 							

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
Other Birds	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> • Avian TB • ChlamydoPhillia • Yersiniosis 							
	Enteric Parasites	major cause of debility	M	H	TREAT	Y	Faeces	Highly recommended
	Enteric bacteria (eg salmonella)	causes disease AND zoonotic	M	M	N	Y	Faeces / cloacal swab	Highly recommended
	Psittacosis (ChlamydoPhillia psittaccae)	Can cause mortality and infertility. common in some wild bird species. Zoonotic. Balai approval will be revoked if positive	M	M/L	Y (TREAT)	y	Blood, faeces	Highly recommended

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LOWER VERTEBRATES AND INVERTEBRATES

ALL ANIMALS SHOULD HAVE:

- Medical history sent a minimum one week prior to export
- Declaration of presence or absence of declaration diseases
- Prophylactic treatments as recommended
- A physical examination – including notification of findings to receiving collection

IT IS HIGHLY RECOMMENDED THAT ALL ANIMALS SHOULD HAVE:

- Tests for the diseases of concern indicated

Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
Lizards	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> • Mortality rate and main causes death in previous year. • OPMV • Cryptosporidiosis 							
	Endoparasites	Can be cause of debility	M	M	N Treat	Fresh faecal examination + floatation	Fresh faeces	3 tests one week apart.
	Paramyxovirus	Has been known to cause death in Rhinoceros iguanas	?M	L	?Y	Blood serology	Serum	Min test twice at 2 month intervals. Test currently available in UK can be difficult to interpret. Declare history and OPMV status of collection. Particularly important if receiving collection is negative see under snakes.
	Cryptosporidiosis	Can be cause of debility Can be a problem for zoos which intend public contact	M	M	Y	Fresh faecal examination	Faeces, If history in collection ? do stomach wash	Declare history of collection.
Snakes	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> • Mortality rate in previous year • OPMV • IBD 							

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
	<ul style="list-style-type: none"> Cryptosporidiosis Amebiasis 							
	Endoparasites	Can be cause of debility	M	M	N Treat	Fresh faecal examination + floatation	Fresh faeces	3 tests one week apart.
	Paramyxovirus	Has been known to cause peracute mortality in snakes.	?H	?M	Y (dependant on status of receiving collection)	Blood serology (only bird PMV tests available in this country – some cross reactivity but significance unclear). PCR developed but not currently commercially available.	Serum (potentially tracheal and cloacal swabs for PCR)	Min test twice at 2 month intervals Declare history and OPMV status of collection. Particularly important if receiving collection is negative. NB interpretation of PMV1 -7 serology results can be difficult in the absence of history of clinical disease.
	Cryptosporidiosis	Can be cause of debility Can be a problem for zoos which intend public contact	M	L	Y	Fresh faecal examination	Faeces, If history in collection ? do stomach wash	Declare history of collection.
	Boid inclusion body disease	Causes morbidity and mortality	H	?M	Y	Biopsies/Histology	Kidney/tonsil/lung/liver	Declare history of collection. In particular nos of snakes died in last 3 years and nos of these that had pm and histology.
Chelonia	<p>Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)</p> <ul style="list-style-type: none"> Upper respiratory tract diseases (URTD) <p>Other:</p> <ul style="list-style-type: none"> Note: Salmonella is not considered to be a disease of concern as all Chelonia should be considered to be carriers and appropriate hygiene measures should be taken. Cryptosporidia are also not included for the same reason. 							
	Enteric parasites	Potential cause of debility	M	H	N	Faecal parasitology	faeces	3 faecal tests one week apart. Including fresh examination for motile protozoa and Ziel Nielsen fro Cryptosporidium. Note many motile protozoa are normal commensals and required for proper gut function.

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
	Mycoplasma	Important cause of mortality/morbidity	H	M	Y/N dependant on status of receiving collection	PCR	Nasal wash, choanal swab	Declare history of upper respiratory tract disease In collection
	Chelonian herpesvirus	Important cause of mortality/morbidity	H-L dependant on species	M	Y/N dependant on status of receiving institution	PCR	Nasal wash, coanal swab	Declare history of upper respiratory tract disease I collection. Current test unable to differentiate between potentially pathogenic and commensal herpes viruses. Interpret results with caution.
Amphibia	Declaration diseases: (must declare if the following found in this taxonomic group of species in previous 12mths)							
	<ul style="list-style-type: none"> • Chytridiomycosis (previous 2yrs) NB NOW A NOTIFIABLE DISEASE • Rana virus (previous 2yrs) NB NOW A NOTIFIABLE DISEASE 							
	Endoparasites	Can be cause of debility	M-H	H	N Treat	Fresh faecal examination + floatation	Fresh faeces	3 tests one week apart.
	Chytridiomycosis	Major cause of death. Major risk for local amphibian fauna.	H	M	Y	Real Time PCR	Skin swab. Skin from post mortem cases, frozen or fixed in 70% ethanol.	Declare history of collection. And mass mortalities for last two years. Indispensable. Treatment + negative testing prior shipment required if positive.
	Ranavirus	Major cause of death.	H	M	Y	PCR	Tissue samples from post-mortem cases	Declare history of collection. No possibility of testing prior to moving the specimens but should be advised to test pm cases? Clear collection history should be obtained prior to transport?
Fish	<ul style="list-style-type: none"> • Prior to moving, I would ensure that a history of the tank/system/species is sent with a particular reference to parasites and infectious diseases. It would be useful to know if any histology has been done and if so how many post mortems/gills presses/skin scrapes and histology have been done cf the number of mortalities from the system/species/tank. • All fish should enter quarantine and the rare exceptions to this mean that pre movement testing is probably less useful than good history. 							

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Taxonomic group of species	Disease of Concern	Justification	HAZARD (H/M/L)	LIKELY HOOD (H/M/L)	SHOW STOPPER? (Y/N)	Screening Test available	Type of sample required	Notes (eg sampling regime, vaccine available / recommended?)
Aquatic inverts (AR +ST)		<p>No requirement for testing pre-move.</p> <ul style="list-style-type: none"> Tank and tank occupant history needed. Histories of treatments e.g. levamisole for nudibranchs on corals but also e.g. <i>Cryptocaryon irritans</i> in fish in shared water. Quarantine needed by recipients. Awareness of disease in local area e.g. crayfish plague in signal crayfish and potentially UK white-clawed. 						
Terrestrial inverts		<p>In terms of disease control, few infectious diseases are well-described and most apparent outbreaks are simply reflecting husbandry stress Therefore, while quarantine is essential the length of time also cannot be known - needs muse tailor the length of quarantine to individual disease and to the species' lifespan.</p> <ul style="list-style-type: none"> If the supplying collection has had previous problems with a potentially infectious agent this should be checked during the quarantine period - where numbers allow this should be done by culling and post-mortem. Also if numbers allow it may be worth culling a few anyway and preserving in alcohol for future investigation should need arise Sick animals should (where numbers allow) be culled for investigation. Dead animals should be stored - there is often little use in performing PM's on these. However, they may be of use for whole body virology, etc should an "outbreak" then start If faeces can be identified endoparasite checks (esp spiders) may be done pre- / post-move Handling animals - screening for salmonella screening not recommended as it would be unclear what either a positive or negative culture would mean. (Most zoonoses are generally contracted by eating the invertebrate!) 						

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Appendix 6. WAZA CODE OF ETHICS AND ANIMAL WELFARE

(Adopted November 2003, San José, Costa Rica)

Preamble

The continued existence of zoological parks and aquariums depends upon recognition that our profession is based on respect for the dignity of the animals in our care, the people we serve and other members of the international zoo profession. Acceptance of the WAZA World Zoo Conservation Strategy is implicit in involvement in the WAZA.

Whilst recognising that each region may have formulated its own code of ethics, and a code of animal welfare, the WAZA will strive to develop an ethical tradition which is strong and which will form the basis of a standard of conduct for our profession. Members will deal with each other to the highest standard of ethical conduct.

Basic principles for the guidance of all members of the World Association of Zoos and Aquariums:

- (i) Assisting in achieving the conservation and survival of species must be the aim of all members of the profession. Any actions taken in relation to an individual animal, e.g. euthanasia or contraception, must be undertaken with this higher ideal of species survival in mind, but the welfare of the individual animal should not be compromised.
- (ii) Promote the interests of wildlife conservation, biodiversity and animal welfare to colleagues and to society at large.
- (iii) Co-operate with the wider conservation community including wildlife agencies, conservation organisations and research institutions to assist in maintaining global biodiversity.
- (iv) Co-operate with governments and other appropriate bodies to improve standards of animal welfare and ensure the welfare of all animals in our care.
- (v) Encourage research and dissemination of achievements and results in appropriate publications and forums.
- (vi) Deal fairly with members in the dissemination of professional information and advice.
- (vii) Promote public education programs and cultural recreational activities of zoos and aquariums.
- (viii) Work progressively towards achieving all professional guidelines established by the WAZA.

At all times members will act in accordance with all local, national and international law and will strive for the highest standards of operation in all areas including the following:

1. Animal Welfare

Whilst recognising the variation in culture and customs within which the WAZA operates, it is incumbent upon all members to exercise the highest standards of animal welfare and to encourage these standards in others. Training staff to the highest level possible represents one method of ensuring this aim.

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Members of WAZA will ensure that all animals in their care are treated with the utmost care and their welfare should be paramount all times. At all times, any legislated codes for animal welfare should be regarded as minimum standards. Appropriate animal husbandry practices must be in place and sound veterinary care available. When an animal has no reasonable quality of life, it should be euthanased quickly and without suffering.

2. Use of Zoo and Aquarium Based Animals

Where "wild" animals are used in presentations, these presentations must:-

- (a) deliver a sound conservation message, or be of other educational value,
- (b) focus on natural behaviour,
- (c) not demean or trivialise the animal in any way.

If there is any indication that the welfare of the animal is being compromised, the presentation should be brought to a conclusion.

When not being used for presentations, the "off-limit" areas must allow the animal sufficient space to express natural behaviour and should contain adequate items for behavioural enrichment.

While the code focuses on zoos and aquarium based "wild" animals, the welfare of domestic animals, e.g., sheep, goats, horses, etc., in, e.g, petting zoos should not be compromised.

3. Exhibit Standards

All exhibits must be of such size and volume as to allow the animal to express its natural behaviours. Enclosures must contain sufficient material to allow behavioural enrichment and allow the animal to express natural behaviours. The animals should have areas to which they may retreat and separate facilities should be available to allow separation of animals where necessary, e.g., cubbing dens. At all times animals should be protected from conditions detrimental to their well-being and the appropriate husbandry standards adhered to.

4. Acquisition of Animals

All members will endeavour to ensure that the source of animals is confined to those born in human care and this will be best achieved by direct zoo to zoo conduct. The advice of the appropriate Species Co-ordinator should be sought before acquiring animals. This will not preclude the receipt of animals resulting from confiscation or rescues. It is recognised that, from time to time, there is a legitimate need for conservation breeding programs, education programs or basic biological studies, to obtain animals from the wild. Members must be confident that such acquisitions will not have a deleterious effect upon the wild population.

5. Transfer of Animals

Members will ensure institutions receiving animals have appropriate facilities to hold the animals and skilled staff who are capable of maintaining the same high standard of husbandry and welfare as required of WAZA

members. All animals being transferred will be accompanied by appropriate records with details of health, diet, reproductive and genetic status and behavioural characteristics having been disclosed at the commencement of negotiations. These records will allow the receiving institution to make appropriate decisions regarding the future management of the animal. All animal transfers should conform to the international standards and laws applying to the particular species. Where appropriate, animals should be accompanied by qualified staff.

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6. Contraception

Contraception may be used wherever there is a need for reasons of population management. The possible side effects of both surgical and chemical contraception, as well as the negative impact on behaviour, should be considered before the final decision to implement contraception is made. (last sentence deleted)

7. Euthanasia

When all options have been investigated and the decision is taken that it is necessary to euthanase an animal, care will be taken to ensure it is carried out in a manner that ensures a quick death without suffering. Euthanasia may be controlled by local customs and laws but should always be used in preference to keeping an animal alive under conditions which do not allow it to experience an appropriate quality of life. Whenever possible a post-mortem examination should be performed and biological material preserved for research and gene conservation.

8. Mutilation

Mutilation of any animal for cosmetic purpose, or to change the physical appearance of the animal, is not acceptable. Pinioning of birds for educational or management purposes should only be undertaken when no other form of restraint is feasible and marking animals for identification should always be carried out under professional supervision, in a way that minimises suffering.

9. Research Using Zoo Based Animals

All zoos should be actively involved in appropriate research and other scientific activities regarding their animals and distribute the results to colleagues. Appropriate areas of research include exhibit design, observations, welfare, behaviour, management practices, nutrition, animal husbandry, veterinary procedures and technology, assisted breeding techniques, biological conservation and cryopreservation of eggs and sperm. Each zoo undertaking such research should have a properly constituted research committee and should have all procedures approved by a properly constituted ethics committee.

Invasive procedures designed to assist in medical research are not to be performed on zoo animals however the opportunistic collection of tissues during routine procedures and collection of material from cadavers will, in most cases, be appropriate.

The well-being of the individual animal and the preservation of the species and biological diversity should be paramount and uppermost in mind when deciding upon the appropriateness of research to be undertaken.

10. Release-to-the-Wild Programmes

All release-to-the wild programmes must be conducted in accordance with the IUCN/SSC/Reintroduction Specialist Group guidelines for reintroduction.

No release-to-the-wild program shall be undertaken without the animals having undergone a thorough veterinary examination to assess their fitness for such release and that their welfare post-release is reasonably safeguarded. Following release, a thorough monitoring program should be established and maintained.

11. Deaths of Animals Whilst in Care

Unless there are sound reasons not to do so, each animal which dies in captivity, or during a release to the wild program, should undergo post-mortem examination and have a cause of death ascertained.

12. External Wild Animal Welfare Issues

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While this code of practice is designed for animals held within Zoos, Aquariums, Wildlife Parks, Sanctuaries, etc., WAZA abhors and condemns ill-treatment and cruelty to any animals and should have an opinion on welfare issues for wild animals external to its membership.

WAZA requires that:

- The taking of animals and other natural resources from the wild must be sustainable and in compliance with national and international law and conform with the appropriate IUCN policy.
- Any international trade in wild animals and animal products must be in compliance with CITES and the national legislation of the countries involved.

WAZA opposes:

- Illegal and unsustainable taking of animals and other natural resources from the wild, e.g. for bush meat, corals, fur or skin, traditional medicine, timber production.
- Illegal trade in wild animals and wild animal products.
- Cruel and non-selective methods of taking animals from the wild.
- Collecting for, or stocking of animal exhibits, in particular aquariums, with the expectation of high mortality.
- The use, or supply of animals for “canned hunting”, i.e. shooting animals in confined spaces, or when semi tranquilised or restrained.
- Keeping and transporting of animals under inadequate conditions, e.g., the keeping of bears in confinement for extraction of bile, dancing bears, roadside zoos or circuses / entertainment.

WAZA and its members should make all efforts in their power to encourage substandard zoos and aquariums to improve and reach appropriate standards. If it is clear that the funding or the will to improve is not there, WAZA would support the closure of such zoos and aquariums.

This document was prepared on the basis of the 1999 Code of Ethics and the 2002 Code of Animal Welfare. It was adopted at the Closed Administrative Session of the 58th Annual Meeting, held on 19th November 2003 at San José, Costa Rica.

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Appendix 7. WAZA GUIDELINES ON THE ACCEPTANCE OF SEIZED OR CONFISCATED ANIMALS

Introduction

1. Live wild animals are seized and confiscated by local, regional and national authorities for a variety of reasons. After seizure, the authorities must ensure that the animals are temporarily placed at a facility where they are housed, fed and cared for according to animal welfare requirements. By the subsequent act of confiscation, the authorities become the owners of the animals and have to dispose of them in a responsible, timely and efficient manner, taking into account practical, legal, animal welfare and conservation aspects.
2. The authorities are assumed to take into account the following guidelines when disposing of confiscated animals:
 - a. the CITES Guidelines for the Disposal of Confiscated Live Specimens of Species included in the Appendices (Resolution Conf. 10. 7, adopted at the 10th Meeting of the Conference of the Parties, Harare (Zimbabwe), 9 to 20 June 1997);
 - b. the IUCN Guidelines for the Placement of Confiscated Animals (approved by the 51st Meeting of the IUCN Council, Gland, Switzerland, February 2000).
Both Guidelines refer to zoos and aquariums as suitable recipients of confiscated animals. They recognise, however, that zoos and aquariums generally cannot accommodate large numbers of animals that become available through confiscations and that, in particular for species with lower conservation value, the authorities may also have to explore other options, such as rescue centres, life-time care facilities, specialist societies, humane societies, commercial captive breeders, or research institutions.
Further guidance is provided to the authorities by
 - c. the IUCN Guidelines for Re-introductions (approved by the 41st Meeting of the IUCN Council, Gland, Switzerland, May 1995); and
 - d. the IUCN Guidelines for the Prevention of Biodiversity Loss Caused by Alien Invasive Species (approved by the 51st Meeting of the IUCN Council, Gland, Switzerland, February 2000).

Acceptance of seized animals

3. Whenever possible, zoos and aquariums should support the efforts of their authorities by accepting to temporarily house, feed and care for seized animals. Institutions accepting such animals may request that their expenses will be reimbursed. It is strongly recommended that arrangements be made under which the costs will be charged to the confiscating authority rather than directly to the importer or owner of the animals.

Advice to authorities regarding placement of animals

4. When confiscating animals, the authorities will have to take the basic decision whether the animals should
 - a. be returned to the wild;
 - b. be maintained in human care for the remainder of their natural lives;
 - c. be euthanised.
5. To facilitate this basic decision, both the CITES and IUCN Guidelines contain decision trees. WAZA and its association members will not interfere with this stage of the decision making process. Individual zoos and aquariums will also refrain from influencing the authorities, unless they are (part of) the CITES Scientific Authority or belong to another government established consultative body and are approached by the authorities in that capacity.

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Acceptance of confiscated animals for permanent keeping

6. Zoos and aquariums will accept confiscated animals only if they have the necessary expertise and can ensure appropriate care and accommodation of the animals in the long term.
7. The animals may be accepted under a permanent loan agreement or as donations. A permanent loan agreement should also define the ownership of the offspring.
8. While the receiving institutions may pay for transportation costs, they should refrain from buying the animals.
9. Zoos and aquariums accepting animals will do so only if the transaction will not result in any benefits to the person or institution from which the animals were confiscated.
10. If the animals belong to a species for which a coordinated regional conservation breeding programme exists, they should be integrated into that programme, if appropriate.

Acceptance of confiscated animals for returning them to the wild

11. If zoos or aquariums are requested by the confiscating authority to accept animals for returning them to the wild, they will accept only if the requirements of the IUCN Guidelines for Re-introductions are met. They will make sure that, during the whole process, these guidelines will be fully respected.

Creating awareness and fundraising for conservation

12. Zoos and aquariums having confiscated animals on display should take the opportunity to inform the public about the reason, which led to the confiscation. In particular, they should make the public aware of the threats unsustainable and illegal trade poses to wild species and of the role CITES plays in combating such trade.
13. Efforts should be made to raise funds for supporting in situ projects for the species concerned, especially in the case of high profile species, such as primates, large carnivores, elephants, rhinos, parrots, or marine turtles etc.

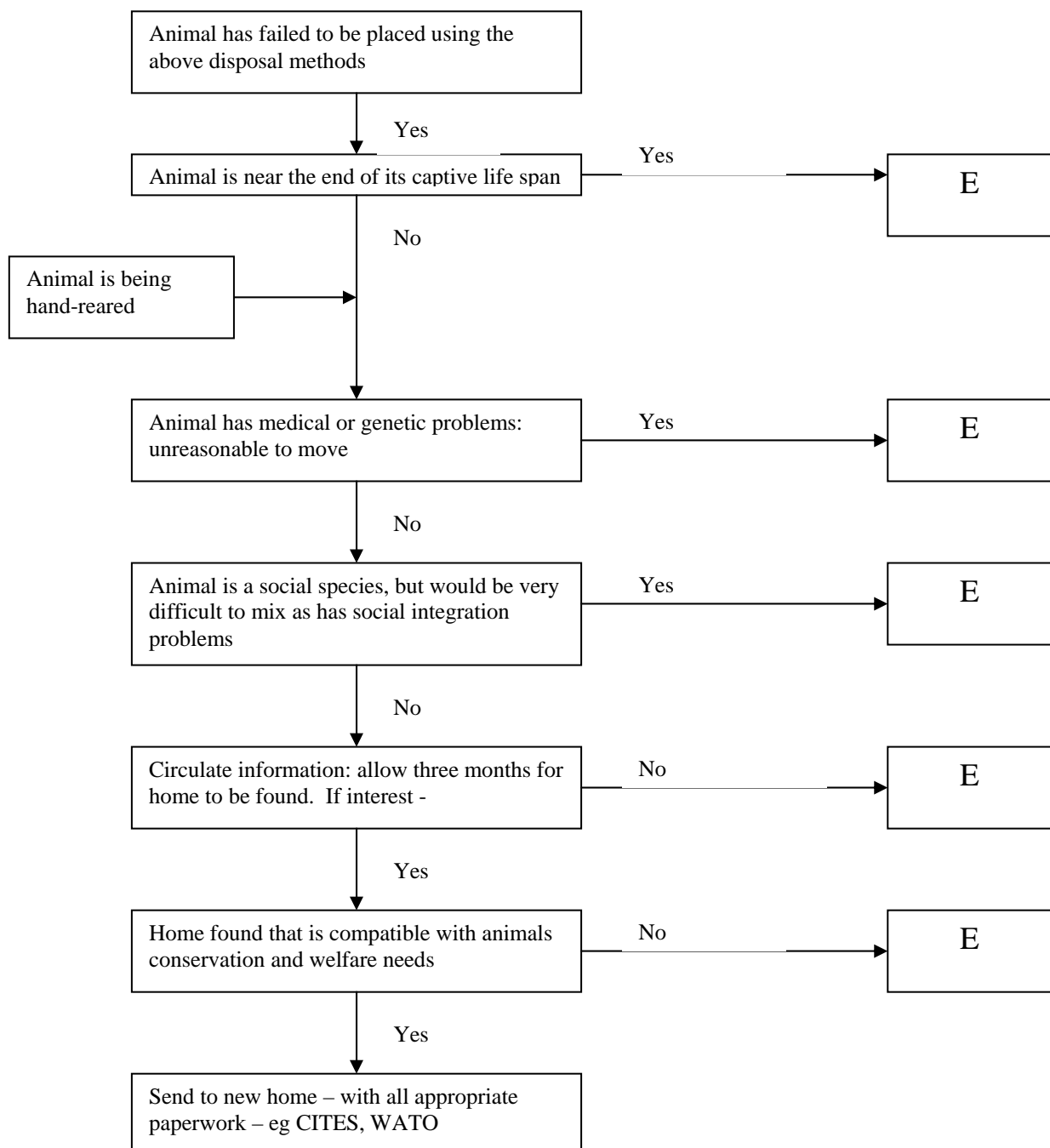
Adopted at the WAZA Plenary Session of 20 November 2003 – 58th Annual Meeting, held at San José, Costa Rica, 2003.

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Appendix 8: Decision Tree (Defra, 2003) on the placement of surplus stock and the decision to cull.

- If after contacting organisations such as BIAZA , the British Association of Leisure Parks, Piers and Attractions, the National Farm Attraction Network; the RSPCA; and European Association of Zoos and Aquaria or other international bodies no home can be found for the animal that is compatible with its conservation and welfare needs;
- The Taxon Advisory Groups (contacted through BIAZA/EAZA) decide the animal cannot be used in a managed programme;
- The owner of the animal (where it was loaned to the zoo) cannot or will not find a home for it or consents to the disposal

The animal should enter the flow chart to decide whether or not euthanasia (E) is an appropriate option



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Appendix 9. IUCN TECHNICAL GUIDELINES ON THE MANAGEMENT OF EX SITU POPULATIONS FOR CONSERVATION

Approved at the 14th Meeting of the Programme Committee of Council, Gland Switzerland, 10 December 2002

PREAMBLE

IUCN affirms that a goal of conservation is the maintenance of existing genetic diversity and viable populations of all taxa in the wild in order to maintain biological interactions, ecological processes and function. Conservation managers and decision-makers should adopt a realistic and integrated approach to conservation implementation. The threats to biodiversity in situ continue to expand, and taxa have to survive in increasingly human-modified environments. Threats, which include habitat loss, climate change, unsustainable use, and invasive and pathogenic organisms, can be difficult to control. The reality of the current situation is that it will not be possible to ensure the survival of an increasing number of threatened taxa without effectively using a diverse range of complementary conservation approaches and techniques including, for some taxa, increasing the role and practical use of ex situ techniques.

If the decision to bring a taxon under ex situ management is left until extinction is imminent, it is frequently too late to effectively implement, thus risking permanent loss of the taxon. Moreover, ex situ conservation should be considered as a tool to ensure the survival of the wild population. Ex situ management should be considered only as an alternative to the imperative of in situ management in exceptional circumstances, and effective integration between in situ and ex situ approaches should be sought wherever possible.

The decision to implement an ex situ conservation programme as part of a formalised conservation management or recovery plan and the specific design of and prescription for such an ex situ programme will depend on the taxon's circumstances and conservation needs. A taxon-specific conservation plan may involve a range of ex situ objectives, including short-, medium- and long-term maintenance of ex situ stocks. This can utilise a variety of techniques including reproduction propagation, germplasm banking, applied research, reinforcement of existing populations and re-introduction into the wild or controlled environments. The objectives and overall purpose should be clearly stated and agreed among organisations participating in the programme, and other relevant stakeholders including landowners and users of the taxon involved. In order to maximise their full potential in conservation, ex situ facilities and their co-operative networks should adopt the guidelines defined by the Convention on Biological Diversity (CBD), the International Agenda for Botanic Gardens in Conservation, Center for Plant Conservation and the World Zoo Conservation Strategy, along with other guidelines, strategies, and relevant legislative requirements at national and regional levels. IUCN recognizes the considerable set of resources committed worldwide to ex situ conservation by the world's zoological and botanical gardens, gene banks and other ex situ facilities. The effective utilisation of these resources represents an essential component of conservation strategies at all levels.

VISION

To maintain present biodiversity levels through all available and effective means including, where appropriate, ex situ propagation, translocation and other ex situ methodologies.

GOAL

Those responsible for managing ex situ plant and animal populations and facilities will use all resources and means at their disposal to maximise the conservation and utilitarian values of these populations, including: 1)

increasing public and political awareness and understanding of important conservation issues and the significance of extinction; 2) co-ordinated genetic and demographic population management of threatened taxa; 3) re-introduction and support to wild populations; 4) habitat restoration and management; 5) long-term

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gene and biomaterial banking; 6) institutional strengthening and professional capacity building; 7) appropriate benefit sharing; 8) research on biological and ecological questions relevant to in situ conservation; and 9) fundraising to support all of the above. *Ex situ* agencies and institutions must follow national and international obligations with regard to access and benefit sharing (as outlined in the CBD) and other legally binding instruments such as CITES, to ensure full collaboration with all range States. Priority should be given to the *ex situ* management of threatened taxa (according to the latest IUCN Red List Categories) and threatened populations of economic or social/cultural importance. *Ex situ* programmes are often best situated close to or within the ecogeographic range of the target taxa and where possible within the range State. Nevertheless a role for international and extra regional support for *ex situ* conservation is also recognised. The option of locating the *ex situ* programme outside the taxa's natural range should be considered if the taxa is threatened by natural catastrophes, political and social disruptions, or if further germplasm banking, propagation, research, isolation or reintroduction facilities are required and cannot be feasibly established. In all cases, *ex situ* populations should be managed in ways that minimize the loss of capacity for expression of natural behaviours and loss of ability to later again thrive in natural habitats.

TECHNICAL GUIDELINES

The basis for responsible *ex situ* population management in support of conservation is founded on benefits for both threatened taxa and associated habitats.

- The primary objective of maintaining *ex situ* populations is to help support the conservation of a threatened taxon, its genetic diversity, and its habitat. *Ex situ* programmes should give added value to other complementary programmes for conservation.

Although there will be taxa-specific exceptions due to unique life histories, the decision to initiate *ex situ* programmes should be based on one or more of the appropriate IUCN Red List Criteria, including:

1. When the taxa/population is prone to effects of human activities or stochastic events or
 2. When the taxa/population is likely to become Critically Endangered, Extinct in the Wild, or Extinct in a very short time. Additional criteria may need to be considered in some cases where taxa or populations of cultural importance, and significant economic or scientific importance, are threatened. All Critically Endangered and Extinct in the Wild taxa should be subject to *ex situ* management to ensure recovery of wild populations.
- *Ex situ* conservation should be initiated only when an understanding of the target taxon's biology and *ex situ* management and storage needs are at a level where there is a reasonable probability that successful enhancement of species conservation can be achieved; or where the development of such protocols could be achieved within the time frame of the taxon's required conservation management, ideally before the taxa becomes threatened in the wild. *Ex situ* institutions are strongly urged to develop *ex situ* protocols prior to any forthcoming *ex situ* management. Consideration must be given to institutional viability before embarking on a long term *ex situ* project.
 - For those threatened taxa for which husbandry and/or cultivation protocols do not exist, surrogates of closely related taxa can serve important functions, for example in research and the development of protocols, conservation biology research, staff training, public education and fundraising.

While some *ex situ* populations may have been established prior to the ratification of the CBD, all *ex situ* and *in situ* populations should be managed in an integrated, multidisciplinary manner, and where possible, in accordance with the principles and provisions of the CBD.

- Extreme and desperate situations, where taxa/populations are in imminent risk of extinction, must be dealt with on an emergency basis. This action must be implemented with the full consent and support of the range State.

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- All ex situ populations must be managed so as to reduce risk of loss through natural catastrophe, disease or political upheaval. Safeguards include effective quarantine procedures, disease and pathogen monitoring, and duplication of stored germplasm samples in different locations and provision of emergency power supplies to support collection needs (e.g. climate control for long term germplasm repositories).
- All ex situ populations should be managed so as to reduce the risk of invasive escape from propagation, display and research facilities. Taxa should be assessed as to their invasive potential and appropriate controls taken to avoid escape and subsequent naturalisation.
- The management of ex situ populations must minimise any deleterious effects of ex situ management, such as loss of genetic diversity, artificial selection, pathogen transfer and hybridisation, in the interest of maintaining the genetic integrity and viability of such material. Particular attention should be paid to initial sampling techniques, which should be designed to capture as much wild genetic variability as practicable. Ex situ practitioners should adhere to, and further develop, any taxon- or region-specific record keeping and genetic management guidelines produced by ex situ management agencies.
- Those responsible for managing ex situ populations and facilities should seek both to increase public awareness, concern and support for biodiversity, and to support the implementation of conservation management, through education, fundraising and professional capacity building programmes, and by supporting direct action in situ.
- Where appropriate, data and the results of research derived from ex situ collections and ex situ methodologies should be made freely available to ongoing in-country management programmes concerned with supporting conservation of in situ populations, their habitats, and the ecosystems and landscapes in which they occur .

NB. Ex situ conservation is defined here, as in the CBD, as "the conservation of components of biological diversity outside their natural habitats". Ex situ collections include whole plant or animal collections, zoological parks and botanic gardens, wildlife research facilities, and germplasm collections of wild and domesticated taxa (zygotes, gametes and somatic tissue).

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Appendix 10. APPROVED CERTIFIED SUSTAINABLE SOURCES

There are number of features unique to marine and some freshwater species that challenge the need to breed every species in captivity. There are also problems associated with the breeding of some marine species which include (Thoney et al, 2003):

- lack of broadly applicable husbandry and veterinary protocols for most major group
- difficulty of satisfying their ecological, behavioural, physiological and nutritional need
- high taxonomic diversity in marine animals and methods of reproduction
- rudimentary state of larval rearing and live food culture techniques.

It is therefore likely that the harvesting of marine species will be necessary to fulfil the needs of public aquariums. Also, in many cases, a sustainable export fishery provides the economic underpinning for community-based conservation programmes that help to preserve the environment from which the fish are collected. Fish harvested from the wild should be collected taking into account:

- human and environmentally non-destructive methods of capture
- harvesting to ensure ecologically sustainability of wild populations

The Marine Aquarium Council (MAC at www.aquariumcouncil.org) is an independent international not-for-profit organisation that brings marine aquarium animal collectors, exporters, importers and retailers together with aquarium keepers, public aquarium, conservation organisations and government agencies. Its Mission is to conserve marine environments and ecosystems by creating standards and certification for those engaged in the collection and care of ornamental marine life from reef to aquarium. It promotes the sustainable use of coral reefs and other marine ecosystems through the responsible collection of ornamental marine life.

Therefore all acquisitions should either be from captive bred stocks (further details in Thoney et al, 2003), or sustainable stocks from the wild, ideally with MAC certification.

Similarly, some terrestrial invertebrates and freshwater fish are also available from sustainable wild sources and there are nationally certified programmes.

Care needs to be taken when acquiring these species that the source is up to scrutiny.

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Appendix 11. WAZA GUIDELINES ON ANIMAL TRANSFERS BETWEEN REGIONS

Background

WAZA recognises that the majority of movements of animals between regions are carefully considered and mutually beneficial. However, the WAZA notes that, in the past, some animal transactions between regions have resulted in:

- *The removal of key animals from coordinated programs in the sending region, thereby disrupting local programs;*
- *The dispersal to another region of animals genetically surplus to the receiving region, to the detriment of the local program.*

WAZA aims to support the development and maintenance of coordinated programs to manage ex situ animal populations for their long-term sustainability. Further, WAZA promotes the principle of mutual support amongst regional associations for regional species management structures.

Accordingly, WAZA urges all regional associations and program coordinators to follow the guidelines outlined below. The guidelines outline WAZA's view on the responsibilities of sending and receiving institutions and species coordinators in the respective regions.

Guidelines for Animal Transfers between Regions

Prior to the transfer of an animal from one region¹ to another:

Both **sending** and **receiving** institutions are responsible for ensuring:

- That the transfer is endorsed by the coordinator of the relevant species management program² operating in their own region, where such a program exists;
- That the proposed transaction is not counter to recommendations made by the relevant advisory body³ in their own region (for example, a Taxon Advisory Group);
- That the counterpart institution has confirmed the same for its own region.

Prior to endorsing the transfer of an animal out of or into a species management program:

The **coordinator** of the species management program is responsible for determining:

- That the transfer of the animal is not detrimental to the species management program;
- That the transfer of the animal is endorsed by the coordinator of the relevant species management program in the other region, where such a program exists.

¹ A 'region' is a geographic area represented by a WAZA-recognised regional zoo and aquarium association.

² A species management program is a program for the coordinated management of the taxon across the relevant region, endorsed by the relevant regional association.

³ An advisory body is one run under the auspices of, or endorsed by, the relevant regional association.

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Practical implications of the Inter-regional Acquisition & Disposition Policy

Institution in Region A – sender	Institution in Region B - receiver
Scenario 1	
No program	Program
<i>Sending institution:</i> <ul style="list-style-type: none"> • checks with relevant TAG, RCP, Association that the move is not contrary to regionally agreed strategy; • seeks assurance from receiving institution that the transfer is endorsed by program in receiving region. 	<i>Receiving institution:</i> <ul style="list-style-type: none"> • seeks endorsement from program coordinator in receiving region.
Scenario 2	
Program	Program
<i>Sending institution:</i> <ul style="list-style-type: none"> • seeks endorsement from program coordinator in sending region; • seeks assurance that receiving institution has done same. <i>Program coordinator in sending region:</i> <ul style="list-style-type: none"> • informs both sending institution and program coordinator in receiving region of endorsement of the transfer. 	<i>Receiving institution:</i> <ul style="list-style-type: none"> • seeks endorsement from program coordinator in receiving region; • seeks assurance that sending institution has done same. <i>Program coordinator in receiving region:</i> <ul style="list-style-type: none"> • informs both receiving institution and program coordinator in sending region of endorsement of the transfer.
Scenario 3	
Program	No Program
<i>Sending institution:</i> <ul style="list-style-type: none"> • seeks endorsement from program coordinator in sending region. 	<i>Receiving institution:</i> <ul style="list-style-type: none"> • checks with relevant TAG, RCP, Association that the transfer is not contrary to regionally agreed strategy; • seeks assurance from sending institution that the transfer is endorsed by program in sending region.
Scenario 4	
No Program	No Program
<i>Sending institution:</i> <ul style="list-style-type: none"> • checks with relevant TAG, RCP, Association that the move is not contrary to regionally agreed strategy; • seeks assurance that receiving institution has done same. 	<i>Receiving institution:</i> <ul style="list-style-type: none"> • checks with relevant TAG, RCP, Association that the transfer is not contrary to regionally agreed strategy; • seeks assurance that sending institution has done same.

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